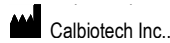


REFERENCES

1. Ashby, J. and Frier, B.: Circulating C-Peptide: Measurement and Clinical Applications. Annals of Clinical Biochemistry, 18:125, 1981
2. Beischer, W.: Proinsulin and C-Peptide in Humans. Hormones in Normal and Abnormal Human Tissues. Volume 3K, Fotherby and Pal, S., ed. (Berlin: Walter DeGruyter). pp. 1-43, 1983
3. Beyer, J., Krause V., Cordes V.: C-Peptide: Its Biogenesis, Structure, Determination and Clinical Significance. Giornale Italiano di Chimica Clinica 4 Supp. 9:22, 1979
4. Bonger, A. and Garcia-Webb, P.: C-Peptide Measurement: Methods and Clinical Utility. CRC Critical Reviews in Clinical Laboratory Sciences, 19:297, 1984.
5. Chevenne D., Ruiz J., Lohmann L., et.al.: Immunoradiometric Assay of Human Intact Proinsulin Applied to Patients with Type 2 Diabetes, Impaired Glucose Tolerance, and Hyperandrogenism. Clinical Chemistry, 40/5:754, 1994
6. Bowsher R. R., Wolny J. D. and Frank B. H.: A Rapid and Sensitive Radioimmunoassay for the Measurement of Proinsulin in Human Serum. Diabetes, 41:1084, 1992
7. Kao P. C., Taylor R. L. and Service F. J.: Proinsulin by Immunochemiluminometric Assay for the Diagnosis of Insulinoma. Journal of Clinical Endocrinology and Metabolism, 78:1048, 1994
8. Dahir F. J., Cook D. B. and Self C. H.: Amplified Enzyme-Linked Immunoassay of Human Proinsulin in Serum (Detection Limit: 0,1 pmol/L). Clinical Chemistry, 38/2:227, 1992

2017-11-08

**Insulin ELISA**

Catalog No. IN374S (96 Tests)

INTENDED USE

The Calbiotech Inc., Insulin ELISA Kit is intended for the quantitative measurement of Insulin in human serum or plasma.

SUMMARY AND EXPLANATION

Insulin is the principal hormone responsible for the control of glucose metabolism. It is synthesized in the β -cells of the islets of Langerhans as the precursor, proinsulin, which is processed to form C-peptide and insulin. Both are secreted in equimolar amounts into the portal circulation. The mature insulin molecule comprises two polypeptide chains, the A chain and B chain (21 and 30 amino acids respectively). The two chains are linked together by two inter-chain disulphide bridges. There is also an intra-chain disulphide bridge in the A chain. Insulin concentrations are severely reduced in insulin-dependent diabetes mellitus (IDDM) and some other conditions such as hypopituitarism. Insulin levels are raised in non-insulin-dependent diabetes mellitus (NIDDM), obesity, insulinoma and some endocrine dysfunctions such as Cushing's syndrome and acromegaly.

PRINCIPLE OF THE TEST

The Calbiotech Inc., Insulin ELISA is based on solid phase sandwich ELISA method. The samples and conjugate reagent (anti-Insulin biotin & HRP) are added to the wells coated with Streptavidin. Insulin in the patient's serum binds to the matched pair Abs, forming a sandwich complex and simultaneously the complex is being immobilized on the plate through streptavidin-biotin interactions. Unbound protein and HRP conjugate are washed off, through a washing step. Upon addition of the substrate, the intensity of color is proportional to the concentration of Insulin in the samples. A standard curve is prepared by relating the color intensity to the concentration of Insulin.

MATERIALS PROVIDED		96 Tests
1.	Microwells coated with Streptavidin	12x8x1
2.	Insulin Standards: 6 vials (ready to use)	0.5 ml
3.	Insulin Conjugate Reagent: 1 bottle (ready to use)	12 ml
4.	TMB Substrate: 1 bottle (ready to use)	12 ml
5.	Stop Solution: 1 bottle (ready to use)	12 ml
6.	20X Wash concentrate: 1 bottle	25 ml

MATERIALS NOT PROVIDED

1. Distilled or deionized water
2. Precision pipettes
3. Disposable pipette tips
4. ELISA reader capable of reading absorbance at 450nm
5. Absorbance paper or paper towel
6. Graph paper

STORAGE AND STABILITY

1. Store the kit at 2 - 8° C.
2. Keep microwells sealed in a dry bag with desiccants.
3. The reagents are stable until expiration of the kit.
4. Do not expose reagent to heat, sun, or strong light.

WARNINGS AND PRECAUTIONS

- For Research Use Only. Not for use in diagnostic procedures.
- For Laboratory use.
Potential biohazardous materials:
The calibrator and controls contain human source components, which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, there is no test method that can offer complete assurance that HIV, Hepatitis B virus or other infectious agents are absent. These reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories" 1984.
- Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
- The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed.
- It is recommended that standards, control and serum samples be run in duplicate.
- Optimal results will be obtained by strict adherence to this protocol. Accurate and precise pipetting, as well as following the exact time and temperature requirements prescribed are essential. Any deviation from this may yield invalid data.

SPECIMEN COLLECTION HANDLING

- Collect blood specimens and separate the serum immediately.
- Specimens may be stored refrigerated at (2-8° C) for 5 days. If storage time exceeds 5 days, store frozen at (-20° C) for up to one month.
- Avoid multiple freeze-thaw cycles.
- Prior to assay, frozen sera should be completely thawed and mixed well.
- Do not use grossly lipemic specimens.

REAGENT PREPARATION

- 20X Wash Buffer Concentrate: Prepare 1X wash buffer by adding the contents of the bottle to 475 ml of distilled water. Store 1X wash buffer at room temperature.

ASSAY PROCEDURE

Prior to assay, allow reagents to stand at room temperature.

Gently mix all reagents before use.

- Place the desired number of coated strips into the holder
- Pipette 25 µl of Insulin standards, control and patient's sera into appropriate wells.
- Add 100µl of Insulin Conjugate Reagent to all wells. Mix well, for 20 seconds.
- Incubate for 60 minutes at room temperature (20-25°C).
- Remove liquid from all wells. Wash wells three times with 300 µl of 1X wash buffer. Blot on absorbent paper towels.
- Add 100 µl of TMB substrate into all wells.
- Incubate for 15 minutes at room temperature.
- Add 50µl of stop solution to all wells. Shake the plate gently to mix the solution.
- Read absorbance on ELISA Reader at 450 nm within 15 minutes after adding the stop solution.

CALCULATION OF RESULTS

The standard curve is constructed as follows:

- Check Insulin standard value on each standard vial. This value might vary from lot to lot. Make sure you check the value on every kit.
- To construct the standard curve, plot the absorbance for the insulin standards (vertical axis) versus the insulin standard concentrations in µIU/ml (horizontal axis) on a linear graph paper. Draw the best curve through the points.
- Read the absorbance for controls and each unknown sample from the curve. Record the value for each control or unknown sample.
- Value above the highest point of the standard are retested after diluting with "0" standard.

Example of Standard Data

	OD 450 nm	Conc. µIU/mL
Std 1	0.007	0
Std 2	0.113	5
Std 3	0.526	25
Std 4	0.914	50
Std 5	1.397	100
Std 6	2.225	300

EXPECTED VALUES

It is strongly recommended that each laboratory should determine its own normal and abnormal values.

In a study conducted with apparently normal healthy adults, using the Insulin ELISA the following values are observed: < 25 µIU/ml.

LIMITATIONS OF THE TEST

- The test results obtained using this kit serve only as an aid to diagnosis and should be interpreted in relation to the patient's history, physical findings and other diagnostic procedures.
- Do not use sodium azide as preservative. Sodium azide inhibits HRP enzyme activities.